

ANTIVIRAL ACTIVITY OF 5-METHYLTHIOMETHYL-2'-DEOXYURIDINE AND OTHER 5-SUBSTITUTED 2'-DEOXYURIDINES

E. DE CLERCQ, J. DESCAMPS, C. L. SCHMIDT and M. P. MERTES

Rega Institute for Medical Research, Katholieke Universiteit Leuven, B-3000 Leuven, Belgium and
Department of Medicinal Chemistry, School of Pharmacy, University of Kansas,
Lawrence, KS 66045, U.S.A.

(Received 10 April 1979; accepted 31 May 1979)

Abstract—Of a series of eight 5-substituted 2'-deoxyuridine (dUrd) derivatives, which were evaluated for their antiviral and antimetabolic activities in primary rabbit kidney or human skin fibroblast cell cultures, five dUrd derivatives, 5-dimethylaminomethyl-dUrd, 5-chloroacetamidomethyl-dUrd, 5-iodoacetamidomethyl-dUrd, 5-pyrrolidinylmethyl-dUrd and 5-N-methylpiperazinylmethyl-dUrd, showed little, if any, activity. The three others, 5-formyl-dUrd, 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd, were found to inhibit the replication of various HSV (herpes simplex virus) strains (whether type 1 or 2) at a concentration of approximately 1–10 µg/ml. The antiviral activity of 5-formyl-dUrd may be accounted for by an inhibition of thymidylate synthetase. The inhibitory effect of 5-formyl-dUrd on HSV multiplication was readily reversed by addition of 2'-deoxythymidine (dTd), and, in analogy with other established thymidylate synthetase inhibitors, 5-formyl-dUrd blocked the incorporation of [2-¹⁴C]dUrd into cellular DNA to a markedly greater extent than the incorporation of [methyl-³H]dTd. Unlike 5-formyl-dUrd, 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd did not preferentially inhibit the incorporation of [2-¹⁴C]dUrd. Antiviral indexes, defined as the ID₅₀ for [2-¹⁴C]dUrd incorporation divided by the ID₅₀ for HSV (type 1, strain KOS) replication, were 0.25, 43 and > 100 for 5-formyl-dUrd, 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd, respectively. The latter two compounds may therefore be considered as rather selective anti-herpes agents.

As recently reviewed by De Clercq and Torrence [1], various 5-substituted 2'-deoxyuridines (dUrd), including 5-fluoro-, 5-chloro-, 5-bromo-, 5-iodo-, 5-nitro-, 5-cyano-, 5-thiocyano-, 5-vinyl-, 5-ethynyl-, 5-allyl-, 5-ethyl-, 5-propyl-, 5-trifluoromethyl-, 5-methylamino-, 5-ethylamino-, 5-methoxymethyl-, 5-mercaptomethyl-, 5-methylmercapto-, 5-hydroxymethyl-, 5-hydroxy-, 5-allyloxy- and 5-propynyloxy-2'-deoxyuridine, have been shown to inhibit the replication of DNA viruses such as herpes simplex virus (HSV) and vaccinia virus. Among these dUrd analogues, 5-propyl-dUrd [2], 5-methylamino-dUrd

[3,4], 5-ethylamino-dUrd [4], 5-methoxymethyl-dUrd [5,6] and 5-propynyloxy-dUrd [7] specifically inhibit the replication of HSV. More recently, two new dUrd derivatives, *E*-5-(2-bromovinyl)-dUrd and *E*-5-(2-iodovinyl)-dUrd, have been added to the growing list of selective antiherpes agents [8–10]. Both compounds were found to inhibit the multiplication of HSV (type 1) at a concentration which was nearly 10,000-fold lower than the concentration required to suppress normal cell growth or metabolism. In marked contrast with the 5-(2-halogenovinyl)derivatives, the 5-(1-halogenovinyl)derivative

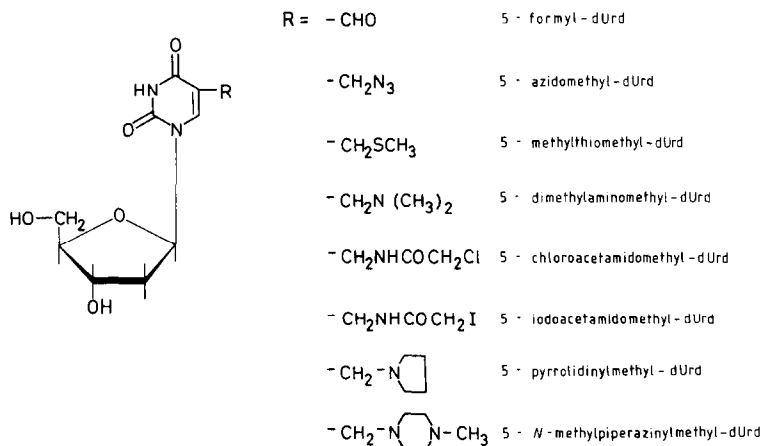


Fig. 1. Structural formulae of 5-substituted 2'-deoxyuridines.

5-(1-chlorovinyl)-dUrd did not exhibit any selectivity in its anti-herpes action [10].

We now report on the antiviral activity of a new series of 5-substituted 2'-deoxyuridines, 5-formyl-, 5-azidomethyl-, 5-methylthiomethyl-, 5-dimethylaminomethyl-, 5-chloroacetamidomethyl-, 5-iodoacetamidomethyl-, 5-pyrrolidinylmethyl- and 5-*N*-methylpiperazinylmethyl-dUrd (Fig. 1). Of this series, only 5-formyl-, 5-azidomethyl- and 5-methylthiomethyl-dUrd were endowed with antiviral properties. For 5-formyl-dUrd the active concentrations coincided closely with the cytotoxic concentrations. 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd were more selective in their antiherpes activity: they inhibited the replication of HSV (whether type 1 or 2) at concentrations which were considerably lower than those that affected normal cell metabolism.

MATERIALS AND METHODS

Compounds. The synthesis and physical characteristics of 5-formyl-dUrd, 5-azidomethyl-dUrd, 5-chloroacetamidomethyl-dUrd, 5-iodoacetamidomethyl-dUrd, 5-dimethylaminomethyl-dUrd, 5-pyrrolidinylmethyl-dUrd and 5-*N*-methylpiperazinylmethyl-dUrd have been described previously [11,12].

The synthesis of 5-methylthiomethyl-dUrd was accomplished by treatment of the 3', 5'-di-*O*-toluoyl ester of 5-chloromethyl-dUrd with methanethiol and base followed by deprotection using potassium carbonate. The spectral characteristics and elemental analyses were as expected.

5-Iodo-dUrd was obtained from Ludco (Brussels, Belgium).

Viruses. Both vaccinia and HSV served as the challenge viruses. For HSV, various laboratory strains or clinical isolates of either type 1 (HSV-1) or type 2 (HSV-2) were employed. The differentiation between HSV-1 and HSV-2 was based upon several criteria: (a) origin (genital or oral), (b) plaque-forming ability in chick embryo cells and (c) virus yields attained in primary rabbit kidney (PRK) cell cultures [13]. The laboratory strains HSV-1 (F), HSV-1 (Mac Intyre) and HSV-2 (G) were obtained from the American Type Culture Collection (Rockville, MD), whereas the laboratory strains HSV-1 (KOS), HSV-2 (LYONS) and HSV-2 (196) were provided by Dr. W. E. Rawls (Baylor University College of Medicine, Houston, TX) through the aid of Dr. J. Desmyter of our Institute. The thymidine kinase deficient (TK⁻) mutant of HSV-1 (B 2006) was kindly given by Dr. Y.-C. Cheng (Roswell Park Memorial Institute, Buffalo, NY). The clinical HSV-1 isolates resulted from either vesicle fluid (VEX 142, VEX 257, VDX 500), mouth swab (VEX 538) or conjunctival swabs (VEX 149, VEO 231, VDO 374, VDX 348). The clinical HSV-2 isolates (VCX 210, VCX 301, VCX 304, VDX 192) originated from cervical swabs. All HSV stocks were prepared in PRK cell cultures.

Inhibition of virus-induced cytopathogenicity (CPE, cytopathic effect). Confluent PRK or HSF (human skin fibroblast) cell cultures in Linbro microtiter trays were inoculated with 100 CCID₅₀ of virus, 1 CCID₅₀ being the virus dose required to infect

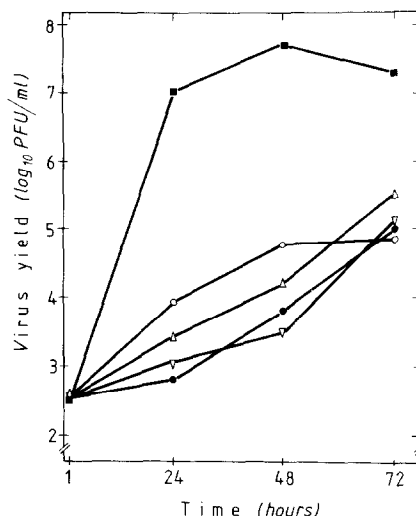


Fig. 2. Effect of 5-formyl-dUrd (∇), 5-azidomethyl-dUrd (\triangle), 5-methylthiomethyl-dUrd (\circ) and 5-iodo-dUrd (\bullet), all at a dose of 10 μ g/ml, on the growth of HSV-1 (strain KOS) in PRK cell cultures. Virus yield was determined at different times after virus infection (as indicated on the abscissa). Control virus yields: \blacksquare . The data represents average values for two separate determinations.

50 per cent of the cell cultures. After 1 hr of virus adsorption, residual virus was removed and the cell cultures were incubated with maintenance medium (Eagle's minimal essential medium supplemented with 3% calf serum) containing varying concentrations of the test compounds (400, 200, 100, . . . μ g/ml). Viral CPE was recorded as soon as it reached completion in the control virus-infected cell structures.

Inhibition of virus growth. Confluent PRK cell monolayers in plastic Petri dishes were inoculated

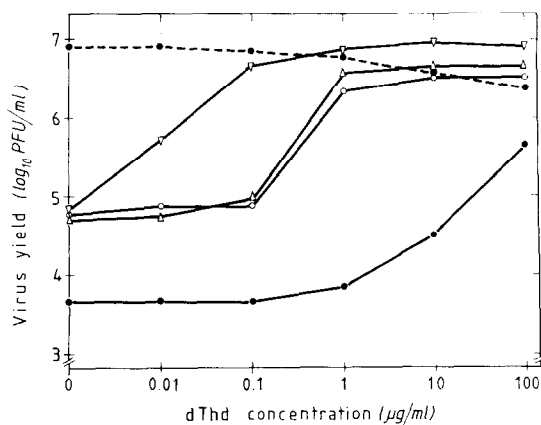


Fig. 3. Reversal of anti-herpes activity of 5-formyl-dUrd (∇), 5-azidomethyl-dUrd (\triangle), 5-methylthiomethyl-dUrd (\circ) and 5-iodo-dUrd (\bullet), all at a dose of 10 μ g/ml, by varying concentrations of dThd (as indicated on the abscissa); dThd alone (--- \bullet ---). Virus yield was determined 24 hr after infection of PRK cell cultures by HSV-1 (strain KOS). The data represents average values for two separate determinations.

with 30,000 PFU (plaque forming units) of HSV-1 (strain KOS) per Petri dish. After 1 hr of virus adsorption, residual virus was removed and the cell cultures were incubated with maintenance medium containing the indicated concentrations of the test compounds (see legends to Figs 2 and 3). Virus yields were determined at the indicated times (see legends to Figs. 2 and 3) by plaque formation in VERO cell cultures.

Inhibition of (methyl-³H)dThd or (2-¹⁴C)dUrd incorporation. The incorporation of [methyl-³H]dThd and [2-¹⁴C]dUrd into PRK cell DNA was monitored as described previously [14]. To this end, PRK cells were seeded in Linbro microwells in the presence of either [methyl-³H]dThd (0.12 μ Ci/0.01 nmole/10⁵ cells) or [2-¹⁴C]dUrd (14 μ Ci/250 nmole/10⁵ cells) and varying concentrations (400, 200, 100, . . . μ g/ml) of the test compounds, and allowed to proliferate for 16 hr at 37° in a humidified, CO₂-controlled atmosphere.

RESULTS AND DISCUSSION

Of the eight 5-substituted dUrd derivatives which were examined for their inhibitory effects on HSV-1 (KOS) and vaccinia virus replication, and [methyl-³H]dThd and [2-¹⁴C]dUrd incorporation into cellular DNA, five compounds, 5-dimethylaminomethyl-dUrd, 5-chloroacetamidomethyl-dUrd, 5-iodoacetamidomethyl-dUrd, 5-pyrrolidinylmethyl-dUrd and 5-N-methylpiperazinylmethyl-dUrd, proved essentially inert as antiviral and antimetabolic agents (Table 1). As 5-iodoacetamidomethyl-dUMP has been previously described as a selective inhibitor of mammalian thymidylate synthetase [15], one may have expected this compound to preferentially inhibit the incorporation of [2-¹⁴C]dUrd [14]. Yet, 5-iodoacetamidomethyl-dUrd did not markedly affect [2-¹⁴C]dUrd incorporation (Table 1). Apparently, 5-iodoacetamidomethyl-dUMP is not sufficiently potent as thymidylate synthetase inhibitor ($K_i/K_m = 3.5$ for calf thymus thymidylate synthetase and 15 for the Ehrlich ascites tumor enzyme [15]) to achieve an appreciable inhibition of [2-¹⁴C]dUrd incorporation *in vivo*.

The relatively low potency of 5-azidomethyl-dUMP as thymidylate synthetase inhibitor ($K_i/K_m = 2.3$ and 1.2 for the calf thymus and Ehrlich ascites tumor enzyme, respectively [16]) may also explain why 5-azidomethyl-dUrd exerted but a weak inhibitory effect on [2-¹⁴C]dUrd incorporation (Table 1). This parallelism may be further extended to 5-methylthiomethyl-dUrd which, on the one hand, did not inhibit [2-¹⁴C]dUrd incorporation into PRK cell DNA (Table 1), and, on the other hand, demonstrated poor activity (in its 5'-monophosphate form) against *Lactobacillus casei*, calf thymus and Ehrlich ascites tumor thymidylate synthetase [$K_i/K_m = 2, 5.8$ and 6, respectively (unpublished observations)].

The results obtained for 5-formyl-dUrd contrasted markedly with those of 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd. 5-formyl-dUrd suppressed [2-¹⁴C]dUrd incorporation at a concentration of 0.5 μ g/ml, that is 100-fold lower than the concentration required to inhibit [methyl-³H]dThd incorpo-

Table 1. Antiviral and antimetabolic activities of 5-methylthiomethyl-dUrd and related 5-substituted 2'-deoxyuridines in PRK and HSF cell cultures

Compound	ID ₅₀ (μ g/ml)					Antiviral index†
	HSV-1 (KOS) PRK	Vaccinia PRK	HSV-1 (KOS) HSF	Vaccinia HSF	(methyl- ³ H)dThd PRK	(2- ¹⁴ C)dUrd PRK
5-Formyl-dUrd	2	0.2	0.4	0.1	50	0.5
5-Azidomethyl-dUrd	0.7	7	1	7	60	30
5-Methylthiomethyl-dUrd	4	200	2	> 200	100	> 400
5-Dimethylaminomethyl-dUrd	> 200	> 200	> 200	> 200	> 400	> 400
5-Chloroacetamidomethyl-dUrd	150	150	150	100	150	> 200
5-Iodoacetamidomethyl-dUrd	> 200	100	> 200	100	100	150
5-Pyrrolidinylmethyl-dUrd	> 200	> 200	> 200	> 200	> 200	> 200
5-N-Methylpiperazinylmethyl-dUrd	> 200	> 200	> 200	> 200	> 200	> 200
5-Iodo-dUrd	0.2	0.2	0.4	0.2	2.5	1.2

* Concentration required to reduce virus-induced CPE, or [methyl-³H]dThd or [2-¹⁴C]dUrd incorporation into DNA of uninfected PRK cells, by 50 per cent.

† Defined as the ratio of the ID₅₀ for [2-¹⁴C]dUrd incorporation to the ID₅₀ for HSV-1 (KOS), both determined in PRK cells.

ation (Table 1). As premised [14], dThd analogues that inhibit dUrd incorporation to a significantly greater extent than dThd incorporation, may be assumed to selectively block thymidylate synthetase *in vivo*. Indeed, thymidylate synthetase is the sole metabolic step that distinguishes the pathways of dUrd and dThd incorporation into DNA dTMP (see Fig. 3 of ref. 14). Thus, 5-formyl-dUrd may be considered as a selective thymidylate synthetase inhibitor *in vivo*. Other examples of dThd analogues that may specifically act at the thymidylate synthetase level *in vivo* include 5-fluoro-dUrd, 5-trifluoromethyl-dUrd, 5-nitro-dUrd, 5-cyano-dUrd, 5-thiocyano-dUrd [14], 5-ethynyl-dUrd, 5-(1-chlorovinyl)-dUrd [10] and, possibly, 5-ethyl-dUrd [14]. That 5-formyl-dUrd may act as an inhibitor of thymidylate synthetase *in vivo* is not unexpected in view of previous studies which point to 5-formyl-dUMP as an extremely potent inhibitor of thymidylate synthetase from *Escherichia coli*, *Lactobacillus casei*, calf thymus and Ehrlich ascites tumor cells [16–19] (K_i/K_m values of 5-formyl-dUMP for these enzymes range from 0.0001 to 0.02 [16–19].)

As noted previously for other selective inhibitors of thymidylate synthetase such as 5-cyano-dUrd [20], 5-thiocyano-dUrd [21], 5-ethynyl-dUrd [10] and 5-nitro-dUrd [14], 5-formyl-dUrd inhibited the replication of both HSV and vaccinia virus (Table 1); and in PRK cells, vaccinia virus was inhibited at a 10-fold lower concentration than HSV-1 (KOS). 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd, two dThd analogues which did not selectively inhibit

thymidylate synthetase *in vivo*, differed from 5-formyl-dUrd in antiviral behavior: these two compounds inhibited HSV replication at a significantly lower concentration than vaccinia virus replication (Table 1). For 5-methylthiomethyl-dUrd, there was even a ≥ 50 -fold difference in the ID_{50} for vaccinia virus and the ID_{50} for HSV-1 (KOS). 5-methylthiomethyl-dUrd may therefore be considered as highly selective in its anti-herpes activity. This selectivity also appeared from toxicity studies which indicated that 5-methylthiomethyl-dUrd did not cause an inhibition of $[2-^{14}C]$ dUrd incorporation (Table 1) or a microscopically detectable alteration of cell morphology (data not shown) at 400 $\mu g/ml$ (100 times the anti-herpes concentration). For 5-azidomethyl-dUrd and 5-formyl-dUrd the minimum concentrations to produce evidence of microscopic cytotoxicity in confluent PRK cell monolayers were 40 and 4 $\mu g/ml$, respectively. Thus, antiviral indexes defined as the minimum dose at which cytotoxicity was observed microscopically divided by the ID_{50} for HSV-1 (KOS) replication were found to be > 100 for 5-methylthiomethyl-dUrd, ~ 50 for 5-azidomethyl-dUrd and 2 for 5-formyl-dUrd. These antiviral indexes correlated well with those based upon the ratio of the ID_{50} for $[2-^{14}C]$ dUrd incorporation to the ID_{50} for HSV-1 (KOS) replication (Table 1). They further point to the lack of specificity in the antiviral action of 5-formyl-dUrd. In accord with previous findings [22], 5-formyl-dUrd inhibited normal cell metabolism and virus multiplication at approximately the same concentration (1–2 $\mu g/ml$).

Table 2. Susceptibility of different HSV-1 and HSV-2 strains to the inhibitory effects of 5-methylthiomethyl-dUrd, 5-azidomethyl-dUrd, 5-formyl-dUrd and 5-iodo-dUrd in PRK cell cultures

Herpes strain	5-methylthiomethyl-dUrd	ID_{50}^* ($\mu g/ml$) 5-azidomethyl-dUrd	5-formyl-dUrd	5-iodo-dUrd
<i>HSV-1 strains</i>				
KOS	4	0.7	2	0.15
F	10	2	2	0.15
Mac Intyre	4	1	0.7	0.15
VEX 142	10	7	2	0.1
VEX 149	20	2	0.4	0.15
VEX 538	20	10	2	0.1
VEO 231	10	10	1	0.15
VEW 257	20	2	1	0.15
VDO 374	2	4	0.4	0.1
VDX 348	40	7	1	0.1
VDX 500	20	2	0.4	0.1
Average for all HSV-1 strains	14.5	4.3	1.2	0.13
<i>HSV-2 strains</i>				
LYONS	4	7	2	0.2
196	4	10	1	0.2
G	4	10	1	0.4
VCX 210	20	1	4	0.3
VCX 301	4	4	1	0.3
VCX 304	4	7	1	0.4
VDX 192	10	4	0.4	0.4
Average for all HSV-2 strains	7.1	6.1	1.5	0.31
HSV-1 TK ⁻ mutant (B 2006)	> 200	≥ 40	2	> 200

* Concentration required to reduce virus-induced CPE by 50 per cent.

The anti-herpes activity of 5-methylthiomethyl-dUrd, 5-azidomethyl-dUrd and 5-formyl-dUrd was not restricted to HSV-1 (KOS). Their antiviral spectrum also encompassed various other laboratory strains of HSV-1 (F, Mac Intyre) and HSV-2 (LYONS, 196, G) as well as clinical HSV-1 and HSV-2 isolates (Table 2). Neither 5-methylthiomethyl-dUrd, 5-azidomethyl-dUrd nor 5-formyl-dUrd discriminated between HSV-1 and HSV-2: they proved equally effective against both types. The standard anti-herpes compound 5-iodo-dUrd, however, was slightly more inhibitory for HSV-1 than for HSV-2 (Table 2). As noted previously [23], HSV type 2 strains tend to be less sensitive to 5-iodo-dUrd than type 1 strains.

While 5-formyl-dUrd inhibited the replication of a dThd kinase deficient (TK^-) mutant of HSV-1 at a concentration of 2 $\mu\text{g/ml}$, thus close to the cytotoxic concentration, 5-methylthiomethyl-dUrd, 5-azidomethyl-dUrd and 5-iodo-dUrd did not effect the replication of the TK^- mutant of HSV-1 (Table 2; for azidomethyl-dUrd the lack of activity could only be established at concentrations up to 40 $\mu\text{g/ml}$, since the compound was cytotoxic from 40 $\mu\text{g/ml}$). The latter results suggest that, as previously shown for several other 5-substituted 2'-deoxyuridines [24,25], 5-methylthiomethyl-dUrd and 5-azidomethyl-dUrd must be phosphorylated by the HSV-induced dThd kinase to exert a selective anti-herpes effect.

That the inhibitory effects of 5-formyl-dUrd, 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd on virus-induced CPE (Table 1) actually reflected an inhibition of virus multiplication was ascertained by measuring the effects of 5-formyl-, 5-azidomethyl- and 5-methylthiomethyl-dUrd on HSV-1 (KOS) growth. When assayed at 100 $\mu\text{g/ml}$, all three compounds as well as 5-iodo-dUrd brought about a significant reduction in virus yield (Fig. 2). For the 24 hr yield reduction in virus titer amounted to 3-4 \log_{10} , and the relative order of (decreasing) activity was 5-iodo-dUrd > 5-formyl-dUrd > 5-azidomethyl-dUrd > 5-methylthiomethyl-dUrd. This is the same order of activity as that obtained for the CPE-inhibition experiments (Table 2).

In agreement with the postulated mode of action of 5-formyl-dUrd, that is an inhibition of thymidylate synthetase (see above), the antiviral activity of 5-formyl-dUrd was readily reversed by dThd: at a concentration 100-fold lower than that of 5-formyl-dUrd, dThd annihilated the inhibitory effect of 5-formyl-dUrd on HSV-1 (KOS) growth (Fig. 3). Similar results have been reported previously for 5-thiocyano-dUrd [21] and 5-nitro-dUrd [14]. These dThd analogues have also been assumed to act as selective inhibitors of thymidylate synthetase [14], and, consequently, they lost their antiviral potency upon addition of extremely low concentrations of dThd (100-fold lower than the concentration at which 5-thiocyano-dUrd and 5-nitro-dUrd were employed) [14, 21]. To abolish the antiviral activity of 5-iodo-dUrd, much higher dThd concentrations were required: even at a 10-fold higher concentration than that of 5-iodo-dUrd, dThd did not completely reverse the inhibitory effect of 5-iodo-dUrd on HSV-1 (KOS) growth (Fig. 3). The relative resistance of the antiviral potency of 5-iodo-dUrd to reversal by dThd,

as noted herein (Fig. 3) and previously [14], would seem compatible with the purported mode of action of 5-iodo-dUrd, that is at the DNA replication or transcription level, thus subsequently to its incorporation into DNA [26].

The inhibitory effects of 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd on herpes virus multiplication could also be reversed by dThd (Fig. 3). This reversal was achieved at a dThd concentration that was intermediate between the dThd concentrations required to abolish the antiviral activities of 5-formyl-dUrd and 5-iodo-dUrd. The fact that the antiviral activity of 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd resisted the reversing effect of dThd more effectively than 5-formyl-dUrd, would strengthen the hypothesis that 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd, unlike 5-formyl-dUrd, do not specifically act at the thymidylate synthetase level. Neither do they seem to act like 5-iodo-dUrd. Otherwise their antiviral activity should have been more resistant to reversal by dThd. 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd may interfere at one or several stages of DNA biosynthesis. Their selective anti-herpes activity seems to depend on the conversion of the nucleoside to the 5'-monophosphate by the virus-induced dThd kinase. Whether the 5'-monophosphate is the active form of 5-methylthiomethyl-dUrd and 5-azidomethyl-dUrd, or whether the compounds must be further converted to the 5'-di- and triphosphate forms to exert their specific anti-herpes action is a matter of further study.

CONCLUSION

Whereas 5-dimethylaminomethyl-dUrd, 5-chloroacetamidomethyl-dUrd, 5-iodoacetamidomethyl-dUrd, 5-pyrrolidinylmethyl-dUrd and 5-N-methylpiperazinylmethyl-dUrd were almost inert as antiviral and antimetabolic agents, 5-formyl-dUrd, 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd exhibited a characteristic biologic activity. 5-Azidomethyl-dUrd and 5-methylthiomethyl-dUrd proved to be specifically active against herpes simplex virus. In this aspect, 5-methylthiomethyl-dUrd is reminiscent of 5-methoxymethyl-dUrd which has also been accredited with a therapeutic index of > 100 [6] and which has recently shown efficacy in the treatment of experimental herpes simplex keratitis in rabbits [27]. In view of its selective activity against herpes simplex virus in cell culture, 5-methylthiomethyl-dUrd could be pursued as an anti-herpes drug.

In marked contrast with 5-azidomethyl-dUrd and 5-methylthiomethyl-dUrd, 5-formyl-dUrd was not specific in its antiviral action. 5-Formyl-dUrd inhibited the replication of both herpes simplex and vaccinia virus, but, at the concentrations that were required to inhibit virus replication, 5-formyl-dUrd also inhibited normal cell metabolism and proliferation [22]. Its antiviral and antimetabolic properties can at least partially be attributed to an inhibition of thymidylate synthetase. Akin to other specific inhibitors of thymidylate synthetase such as 5-fluoro-dUrd and 5-trifluoromethyl-dUrd [28], 5-formyl-dUrd should be pursued as an antitumor agent.

Acknowledgements—This investigation was supported by grants from the Belgian Fonds voor Geneeskundig Wetenschappelijk Onderzoek (Krediet 3.0048.75) and the Geconcerteerde Onderzoeksacties (Conventie n° 76/81-IV). We thank Frieda De Meyer for excellent technical assistance.

REFERENCES

1. E. De Clercq and P. F. Torrence, *J. Carbohydr. Nucleos. Nucleot.* **5**, 187 (1978).
2. E. De Clercq, J. Descamps and D. Shugar, *Antimicrob. Ag. Chemother.* **13**, 545 (1978).
3. M. M. Nemes and M. R. Hilleman, *Proc. Soc. exp. Biol. Med.* **119**, 515 (1965).
4. T. Y. Shen, J. F. McPherson and B. O. Linn, *J. med. Chem.* **9**, 366 (1966).
5. J. B. Meldrum, V. S. Gupta and J. R. Saunders, *Antimicrob. Ag. Chemother.* **6**, 393 (1974).
6. L. A. Babiuk, B. Meldrum, V. S. Gupta and B. T. Rouse, *Antimicrob. Ag. Chemother.* **8**, 643 (1975).
7. P. F. Torrence, J. W. Spencer, A. M. Bobst, J. Descamps and E. De Clercq, *J. med. Chem.* **21**, 228 (1978).
8. R. T. Walker, P. J. Barr, E. De Clercq, J. Descamps, A. S. Jones and P. Serafinowski, *Nucleic Acids Res. (special publication)* **4**, S103 (1978).
9. E. De Clercq, J. Descamps, P. J. Barr, A. S. Jones, P. Serafinowski, R. T. Walker, G.-F. Huang, P. F. Torrence, C. L. Schmidt, M. P. Mertes, T. Kulikowski and D. Shugar, in *Antimetabolites in Biochemistry, Biology and Medicine* (Eds. J. Skoda and P. Langen), Pergamon Press, Oxford p. 275 (1979).
10. E. De Clercq, J. Descamps, P. De Somer, P. J. Barr, A. S. Jones and R. T. Walker, *Proc. natl. Acad. Sci. U.S.A.* **76**, 2947 (1979).
11. A. Kampf, C. J. Pillar, W. J. Woodford and M. P. Mertes, *J. med. Chem.* **19**, 909 (1976).
12. M. S. Edelman, R. L. Barfknecht, R. Huet-Rose, S. Boguslawski and M. P. Mertes, *J. med. Chem.* **20**, 669 (1977).
13. E. De Clercq, J. Descamps, G. Verhelst, R. T. Walker, A. S. Jones, P. F. Torrence and D. Shugar, submitted for publication (1979).
14. E. De Clercq, J. Descamps, G.-F. Huang and P. F. Torrence, *Molec. Pharmac.* **14**, 422 (1978).
15. R. L. Barfknecht, R. A. Huet-Rose, A. Kampf and M. P. Mertes, *J. Am. chem. Soc.* **98**, 5041 (1976).
16. A. Kampf, R. L. Barfknecht, P. J. Shaffer, S. Osaki and M. P. Mertes, *J. med. Chem.* **19**, 903 (1976).
17. D. V. Santi and T. T. Sakai, *Biochem. biophys. Res. Commun.* **42**, 813 (1971).
18. D. V. Santi and T. T. Sakai, *Biochem. biophys. Res. Commun.* **46**, 1320 (1972).
19. Y. Wataya, D. V. Santi and C. Hansch, *J. med. Chem.* **20**, 1469 (1977).
20. P. F. Torrence, B. Bhooshan, J. Descamps and E. De Clercq, *J. med. Chem.* **20**, 974 (1977).
21. E. De Clercq, P. F. Torrence, J. A. Waters and B. Witkop, *Biochem. Pharmac.* **24**, 2171 (1975).
22. P. Langen, S. R. Waschke, K. Waschke, D. Bärwolff, J. Reefschräger, P. Schultz, B. Preussel and C. Lehmann, *Acta biol. med. germ.* **35**, 1625 (1976).
23. P. Collins and D. J. Bauer, *Ann. N.Y. Acad. Sci.* **284**, 49 (1977).
24. Y.-C. Cheng, B. A. Domin, R. A. Sharma and M. Bobek, *Antimicrob. Ag. Chemother.* **10**, 119 (1976).
25. E. De Clercq, E. Krajewska, J. Descamps and P. F. Torrence, *Molec. Pharmac.* **13**, 980 (1977).
26. B. Goz, *Pharmac. Rev.* **29**, 249 (1978).
27. J. B. Meldrum, V. S. Gupta and L. A. Babiuk, *Chemotherapy*, in press (1979).
28. P. V. Danenberg, *Biochim. biophys. Acta.* **473**, 73 (1977).